



MACROLACTONES AND MACROLIDES FROM PLANT ENDOPHYTIC FUNGI, CHEMICAL SCAFFOLDS, BIOLOGICAL ACTIVITIES AND SPECTROSCOPY: A COMPREHENSIVE REVIEW

Ehab Saad El-khayat^{1*}, Mohamed E. Abouelela^{1,3,4} Reda Ahmad Abdelhamid¹ and Mohammad S. Alorainy² and Khaled A. Shaaban^{3,4}

¹Department of Pharmacognosy, Faculty of Pharmacy, Al-Azhar University, Assiut-Branch, Assiut 71524, Egypt

²Department of Pharmacology, College of Medicine, Qassim University, 6655 Buraidah 51542, Saudi Arabia

³Center for Pharmaceutical Research and Innovation, College of Pharmacy, University of Kentucky, Lexington, Kentucky 40536, United States

⁴Department of Pharmaceutical Sciences, College of Pharmacy, University of Kentucky, Lexington, Kentucky 40536, United States

Background: The pandemic of COVID-19 has stressed the exaggerated demand for innovative treatments, prompting the search for new sources. Plant endophytic fungi produce a diverse array of biologically active compounds, including macrolides and macrolactones with varying activities. Aim of the Study: In this review we give an updated overview of natural macrolides and macrololactones from plant endophytes addressing original studies published up to June 2023. Results: Over the preceding ten years, 91 macrolides with 80 novel compounds with cytotoxic, antibacterial, antifungal, and α -glucosidase inhibitory activities. Unfortunately, the number of novel chemicals identified from marine or bacterial endophytes in the same period is substantially lower. Accordingly, further study on plant endophytes, which are critical for drug research and the development of novel medicines, including antitumors, antivirals, antibacterials, and antimalarials, should be conducted. A report of the ¹³C NMR data of several endophytic macrolides are reported as a supplementary according to ring sizes and based on a united numbering built on literature search

Keywords: Macrolides, plant endophytes, biosynthesis, biological activity, ¹³C-NMR

INTRODUCTION

Natural products, particularly those generated by microorganisms or due to their interactions with their hosts, continually offer evidence for their critical role in medication development ¹. Natural product-driven drug development has improved, either directly from natural compounds or through semi-synthetic and synthetic derivatives inspired by natural product prototypes. Over the previous three decades, these compounds have accounted for more than half of all authorized small-molecule

medications². Metabolites from microorganisms, particularly fungi, and their symbiotic interaction products with other or marine species have been terrestrial recruited for natural product research. Endophytic fungi (endophytes) are а remarkable symbiotic microorganism-host interaction that has produced a plethora of new and biologically active metabolites^{3,4}. These metabolites have displayed various biological capabilities, including antibacterial, antifungal, immunosuppressive, antiviral, antimalarial, anti-inflammatory, and anticancer effects^{5,6}.

Received : 22/7/2023 & Accepted : 16/9/2023

^{*}Corresponding author: Ehab Saad El-khayat, E-mail: elkhayat@azhar.edu.eg

Endophytic metabolites are structurally classified into alkaloids, benzopyranones, peptides, macrolides, quinones, steroids, terpenoids, tetralones, xanthones, and others ⁷.

Endophytic fungal macrolides are polyketides with variable-sized macrolactone ring scaffolds that have a wide range of biological actions, including antiparasitic, antimalarial effects^{8,9} antifungal, and Clinically used macrolides are effective against gram-positive and atypical bacteria. Because of their immunostimulant and anti-inflammatory properties, they have been explored as a wide adjuvant treatment for COVID-19. During the COVID-19 pandemic. repurposing of azithromycin, in particular, inhibits proinflammatory cytokine production (including MMPs, TNF- α , IL-6, and IL-8), increases levels of interferons and interferon-stimulated proteins, and reduces viral multiplication and release ^{10,11}. Since the COVID pandemic, repurposing of the existing medications to achieve new therapeutic effects has gained increased attention as a viable technique to accelerate drug development while minimizing experimental costs and time¹².

Accordingly, macrolides have acquired growing importance as a research topic for academic institutions and pharmaceutical research centers, steering efforts to identify novel molecules by chemical modification of existing natural skeletons¹³.

On tracing previous macrolide articles which were published during the last decade, many reviews have been reported concerned with macrolides^{14,15}, for example Lenz and colleagues investigated the structures, sources, mechanism of action, and biological effects of selected macrolides with potential therapeutic uses ¹⁶. In addition to certain papers addressing particular classes of macrolides with their biological activity, and chemical or biological synthesis¹⁷⁻¹⁹. This review provides an updated overview of new macrolides and macrolactones isolated from plant fungal endophytes, with an emphasis on new chemical structure and biological activity, or new biological activity of known unevaluated compounds in the time between 2013 and June 2023. This thorough review will be a helpful resource for natural product and medicinal chemistry researchers, as well as biologists and for drug development and/or drug repurposing.

MACROLIDES AND MACROLACTONES

Macrolactones

Macrolactones are macrocyclic lactones having at least twelve atoms in their core ring structure, they enclude a wide range of natural compounds with variable biological and helpful druglike activities ²². The macrolactones are reported in the following text and **Fig. 2-4** based on their ring sizes.

Ten-membered macrolactones

The 10-membered macrolactones are metabolites derived from marine²¹, pathogenic²² and soil derived fungi²³. They have been reported with various biological activities including cytotoxicity²⁴, antimalarial²⁵ and anti-inflammatory²⁶. The reported 10-membered macrolactones in the last decade are described in this section and in **Fig. 1**.

Colletotriolide 1 was isolated from the leaves of the Chinese mangrove Pandanus amaryllifolius by the endophytic fungus Colletotrichum sp. It inhibited Escherichia coli with a modest IC_{50} of 500 mg/mL²⁷. Four more congeners were recovered from *Phomopsis* sp., an endophyte isolated from Pinus massoniana in south China. They were identified according to IUPAC system as (5S,8S,9R,10R,E)-5,8,9trihydroxy-10-pentyl-3,4,5,8,9,10,-hexahydro-2H-oxecin-2-one 2, (5S,8S,9R,10R,E)-5,8,9trihydroxy-10-nonyl-3,4,5,8,9,10,hexahydro-2H-oxecin-2-one 3, (5R,8S,9R,10S,E)-5,8,9trihvdroxy-10-([*R*]-4-hvdroxyoctyl)-3,4,5,8,9,10-hexahydro-2*H*-oxecin-2-one 4, and (5*S*,6*S*,9*S*,10*R*,*E*)-5,6,9-trihydroxy-10-pentyl-3,4,5,6,9,10-hexahydro-2*H*-oxecin-2-one 5 ²⁸. Compound 2 which acquired the name seimatopolide A was previously isolated from Seimatosporium sp., the endophyte from *Hypericum* perforatum²⁹. Hiep and his colleagues investigated this compound as an agonist of the peroxisome proliferator-activated receptor (PPAR)- γ as a potential therapeutic option for diabetes therapy $(EC_{50} \text{ of } 1.15 \text{ M})^{30}$. The polyhydroxy congener Mangiferaelactone 6 was isolated from the Panama shrub Pestalotiopsis manguiferae, an endophytic fungus associated with Hyptis dilatate. It inhibited the growth of Enterococcus faecalis, Bacillus cereus, Enterococcus cloacae, Listeria monocytogenes, and Proteus mirabilis 31 Botero and co-researchers isolated two new modiolides D. E and A 7-9 from the endophyte Microsphaeropsis arundinis³². Modiolide A 9 was previously reported from the marine fungus Paraphaeosphaeria sp. These compounds revealed weak cytotoxic activities against murine breast (LM3), murine lung (LP07) and human breast (MCF-7) cell lines 33 . Pedra and his group, isolated the possible glioma therapy, Sch-642305 10 from the unidentified endophyte MF31b11 of the Brazilian medicinal plant Achyrocline satureioides. It showed anti-proliferative properties against C6 and U138MG glioma cells, with IC_{50} of 1.1 and 7.6µg/mL, respectively, it promoted apoptosis, decreased cell migration, enhanced antioxidant defense system and suppressed ROS production³⁴. The three (11S) xestodecalactones derivatives xestodecalactones D-F 11-13, were isolated by

Ebrahim and his group from *Corynespora cassiicola*, the endophyte from Chinese mangrove *Laguncularia racemosa* ³³. Zhang and his group have isolated hispidulactone B 14 from the endophyte *Chaetosphaeronema hispidulum* ⁴. Two years later, Zheng and his group have isolated hispidulactone F 15 from the same fungus in 2020, both compounds 14 and 15 inhibited HepG2 cell proliferation in a dose-dependent manner with IC₅₀ 61.05 and 107.69 μ mol/L, respectively³⁵.

Twelve membered macrolctones

Numerous 12-membered macrolactones with various biological properties, including antimicrobial and anti-inflammatory activity, have been isolated from plant endophytes ²⁸. The 12-macrolactones, including derivatives of resorcylic acid and curvularin, are described in this section and in **Fig. 2**.



Fig. 1: Ten membered macrolctones.



Fig. 2: Twelve membered macrolactones.



Fig. 3. Thirteen membered macrolctones.

C-12 macroloides

Investigation of the mangrove endophyte, *Cladosporium cladosporioides* MA-299, afforded 5*R*-hydroxyrecifeiolide 16 and 5*S*-hydroxyrecifeiolide 17 in addition to pandangolide 1 18³⁷. Pandangolide 1 18 was previously isolated from undescribed marine fungus³⁸. Liu and his group have identified 4-hydroxy-12-methyloxacyclododecane-2,5,6-

trione 19 and 12-methyloxacyclo-dodecane-2,5,6-trione 20 from *Cladosprium colocasiae* A801, the endophyte from the Australian shrub *Callistemon viminalis*. A panel of biological evaluation of both compounds including antibacterial against *Staphylococcus aureus*, *Escherichia coli* and MRSA, in addition to cytotoxicity against SF-268, MCF-7, and HepG-2 cell lines, as well as α -glucosidase inhibitory activity, didn't show any positive results⁴¹.

The research group of Wuringege isolated two new pandangolide derivatives from *Cladosporium* sp. IFB3lp-2, the foliar endophyte of *Rhizophora stylosa*, Hainan Island, China. Compounds were identified as methyl-2-((4*R*,6*S*,12*R*)-6-hydroxy-12-methyl-2,5-dioxooxacyclododecan-4-yl)thio)-acetate 21 and (E)-(3*R*,6*S*)-6-Hydroxy-12-methyl-2,5dioxooxacyclododecan-3-yl4,11-

dihydroxydodec-2-enoate 22^{4^2} . The infrequent C-2 sulfur substituted metabolites, thiocladospolides A–D 23-26 were isolated from *C. cladosporioides* MA-299, from the Chinese mangrove *Bruguiera gymnorrhiza*. Compound 23 showed strong inhibition against the aquatic pathogen *Edwardsiella tarda*, while compound 26 showed strong inhibition against *E. ictarda*, both with an MIC of 1 µg/mL. The known pandangolide 3 27 was isolated and revised as well, where the sulfur side chain

been located at C-2 through extensive NMR analysis and in comparison to compound 23 43 .

Investigation of Cladosporium sp. SCNU-F0001 afforded two new lactam macrolide cladospamide A 28, and thiocladospolide E 29 44,45 Thiocladospolides F-J 30-34 were isolated from Cladosporium oxysporum which was isolated from the roots of Avicennia marina ⁴⁵. Further investigation of the Cladosporium endophytes led to isolation of two sulfur-containing macrolides 35 and 36 from Cladosporium cladosporioides MA-299, both compounds were active against the aquatic pathogenic bacteria Edwardsiella tarda and Vibrio anguillarum with MIC range from 2.0 to 4.0 μ g/mL ³⁹, unfortunately these compounds were given the names thiocladospolides F and G, which is exactly the same names of compounds 30 and 31 described by wang in 2020, this nomenclature was misguiding as the same name identifies two different compound structures. Hence, we suggest that compounds 35 and 36 should be characterized as thiocladospolides K and L, instead.

Curvularins

The curvularins are substituted resorcinol fused to the β , γ -positions of a macrocyclic lactone ring. They are produced by several fungal genera as *Aspergillus*⁴⁴, *Alternaria*⁴⁵, *Curvularia*²⁷ and *Penicillium*⁴⁶.

The sulfur-containing curvularins, sumalarins A-C 37-39 were obtained from *Penicillium sumatrense* MA-92 the endophyte associated with the mangrove *Lumnitzera racemose* ⁵⁷. An unusual bicyclo 5/9 ring system C12-macrolide, cladocladosin A 40, was isolated from *Cladosporium cladosporioides* MA-299 from the mangrove *Bruguiera gymnorrhiza*, collected from Hainan Island, China. It showed marked antimicrobial activity against *E. tarda*, *V. anguillarum* and *P. aeruginosa* with MIC of 1, 2 and 4 μ g/mL, respectively⁴⁸.

Resorcylic acid lactones

The resorcylic acid lactones are metabolites having a β -resorcylic acid and a 12 or 14-membered lactone ring with a C-10 methyl substituent^{49,50}. These compounds have wide range of biological activities including antiplasmodial⁵⁰, and cytotoxic⁵¹, in addition to estrogenic and kinase inhibitory activities⁴⁹.

The β -resorcylic acid lactone lasiodiplodin 41. which was initially reported as plant metabolite from Euphorbia splendens 52 and later was purified from the endophyte Lasiodiplodia sp isolated from the Chinese mangrove Acanthus ilicifolius, in addition to (E)-9-etheno-lasiodiplodin 42. Both compounds showed α -glucosidase inhibition activity with IC_{50} 32.5 and 35.9 μ M, respectively⁵³. Later, lasiodiplactone A 43 was isolated from Lasiodiplodia theobromae ZJ-HQ1, the endophyte of the same mangrove plant. The absolute configuration was assigned 15R, 18S, 19S, 21S by comparing as experimental and calculated ECD spectra using time-dependent density-functional theory (TDDFT). Its biological evaluation showed anti-inflammatory activity through inhibition of nitric oxide production in lipopolysaccharide activated RAW264.7 cells with IC₅₀ of 23.5 μ M, and α -glucosidase inhibitory activity with IC₅₀ 29.4 µM⁵⁴.

Thirteen Membered macrolactones

Several 13-membered macrolctones were isolated from several fungal species (Fig. 4), biologically they were evaluated as antimicrobial, anticancer, and chemopreventive agents^{55,56}. According to the National Cancer Institute's in vitro anticancer screening, brefeldin Α 43 proved to have а chemotherapeutic activity. It is an interesting C-13 macrolide with antibiotic, antiviral, cytostatic, antimitotic, and antitumor activities. It was previously isolated from several fungal genera as Alternaria, Ascochyta, Penicillium, Curvularia, Cercospora and Phyllosticta ⁵⁷. Five new brefeldin A congeners, brefeldin A 7-O-acetate 44 and the open ring derivatives brefeldin E1-E5 45-49, were isolated from Penicillium sp., the endophyte of Panax

notoginseng root. These compounds displayed low cytotoxic activities⁵⁸.

The *N*-demethylmelearoride A **50** was isolated from the solid culture of the endophyte *Penicillium brefeldianum* XMK-2 isolated from the rhizome of *Pinellia ternate*. It has moderate cytotoxic activity, against HepG2 cells with IC₅₀ of 36.6 μ mol/L⁵⁹.

MACROLIDES

The term macrolides denotes those possessing 14-,15-, 16- and larger macrolactones rings constitute a family of natural origin with wide range of biological potency 23 . In the following text and in **Fig. 4- 6**, we report the macrolides according to their ring size.

Fourteen membered macrolides

This class of macrolides have variable activities as the phytotoxic seiricurprolide from *Seiridium cupressi*⁶⁰, and zearalenone the mycotoxin with estrogenic activity from the genus *Fusarium*⁶¹, and the cytotoxic aspergillides A–C from *Aspergillus ostianus*,⁶².

Liu and his group have isolated seven new 14-membered macrolides, (13S)pestalotioprolides C-H 51-56 and 7-0methylnigrosporolide 57 (Fig. **4**) from Pestalotiopsis microspore, the endophyte of the mangrove Drepanocarpus lunatus⁶³, in addition to nigrosporide 58 which was previously Nigrospora sphaerica⁶⁴ isolated all compounds were proved to have the 13S configuration by Single-crystal X-ray analysis, Mosher's and TDDFT-ECD experiments. Compounds 52-4 and 57 showed cytotoxic activity against L5178Y murine lymphoma cell line, while compound 53 showed potent activity against human ovarian cancer A2780 cell line with an IC₅₀ of 1.2 μ M⁶⁵. Chen *et al.*, have isolated a C-14 β -resorvelic macrolactone 3-methoxy-lasicicol derivative. 59 from Lasiodiplodia sp. ZJ-HQ the endophyte associated with Acanthus ilicifolius⁵³. It is the methyl derivative of the known lasicicol 60 from Saccharomyces cerevisiae²¹, which was also isolated in this work, X-ray diffraction analysis had confirmed the structure of 60, and the absolute configuration was determined by modified Mosher's experiment. Both compounds showed potent α -glucosidase inhibitory activity ⁵³.



Fig. 4: Fourteen membered macrolides.

Sixteen membered macrolide

16-membered macrolides The is a significant class of macrolides with antibiotic activity. They have been isolated from marine or terrestrial fungi, such as chalcomycin B, from the marine Streptomycete B7064, it exhibited antimicrobial activities against a variety of microorganisms and microalgae⁶⁶. Later on, Juvenimicin C, was isolated from the marine *Micromonospora* sp. CNJ-878⁶⁷. The only natural 16-membered macrolides in clinical use today are josamycin, spiramycin and midecamycin, in addition to the semisynthetic miokamycin, rokitamycin and tildipirosin (first available in 2012). The antiinflammatory and immunomodulatory effects have also been reported for C-16 macrolides, as well as the antimalarial activity of tylosin A and derivatives of desmycosin (tylosin B)⁶⁸. In the following text and in Fig. 5 we report the C-16 plant endophytic macrolides isolated during the last 10 years.

homodimeric C-16 macrolides, Two pyrenophorin 61 and pyrenophorol 62 were isolated from the needle endophyte Lopherdermium nitens of the Canadian Pinus strobus⁶⁹. Compound 61 showed significant antifungal activity against Cronartium ribicola at 5 μ M⁷⁰. McMullin and his group have demonstrated that 62 significantly reduced the biotrophic growth of the pathogen Microbotryum violaceum and Cronartium ribicola at 4 and 5 µM, respectively. Both 61 and 62 have been previously identified from endophytes of Lycium intricatum^{69,71}.

Tricothecene macrolides

The trichothecene macrolides is a varied class of fungal sesquiterpenoids with acyl residue(s) tethered at 4β and/or 15 positions⁷². They are characteristic for species of the genera Fusarium, Myrothecium, Trichoderma, Trichothecium, Cephalosporium, Verticimonosporium, and Stachybotrys⁷³. Many trichothecenes have confirmed anticancer⁷⁴ immunomodulation^{75,}, phytotoxicity⁷⁶, activities⁷⁸. antifungal⁷⁷ and antimalarial According to the carbon skeleton, they are classified into C_{27} vertucarins and C_{29} roridins^{72,78}.

The new cytotoxic roridin-type trichothecene, roritoxin E 63 was isolated from Myrothecium roridum IFB-E091, showed in vitro inhibitory effect against gastric carcinoma SGC-7901 and hepatocarcinoma SMMC-7721 cell lines⁷⁹. The myrothecines D-G 64-6769-72, 16-hydroxymytoxin and 14'-В 68. dehydrovertisporin 69 were obtained from Myrothecium roridum. All compounds showed antiproliferative effect against chronic myeloid leukemia K562, and colorectal carcinoma (SW1116) cell lines⁷². Meanwhile two new roridin-type trichothecenes, myrothecines H and I 70 and 71 were isolated from Paramyrothecium roridum,, both have been reported with high cytotoxic activity against SF-268 and HepG-2 cell lines⁷⁹. Epiroridin acid 72, epiroridin E 73 and mytoxin B 74 were isolated from the liquid culture of Myrothecium roridum A553, the endophyte from the medicinal plant Pogostemon cablin. All compounds were evaluated for their in vitro cytotoxic activities against human glioma, human breast adenocarcinoma (MCF-7), human non-small cell lung cancer (NCI-H460), and human hepatoma (HepG-2)⁸⁰. From the endophyte *M. roridum* IFB-E012. Shen and his group have isolated dihydromyrothecine C 75 as an epimeric mixture of the (14'S) and (14'R)stereoisomers, the (14'S) was more stable. It showed moderate cytotoxicity with IC₅₀ 44.48 µM against human nasopharyngeal carcinoma cell line (KB)⁸¹. The known verrucarin A 76⁸² isolated from the endophyte was Paramyrothecium roridum associated with Morinda officinalis and evaluated as antiproliferative and apoptosis-inducing agent against CaP cells⁸⁰. Nguyen et al., reported a nematocidal activity for this compound at a concentration of 1.88 μ g/mL⁸³.

The 2',3'-epoxymyrothecine A 77 and 13',14'-hydroxymytoxin B 78, in addition to the known mytotoxin A 79 were isolated from the endophyte *Myrothecium roridum* associated

with the Chinese herb *Ajuga decumbens*. It showed potent cytotoxic activity against A549, MCF-7, HepG2, and 7721 cell lines, 79 cell lines. Meanwhile, cell cycle arrest investigations showed that 77, and 79 could induce G1 arrest in HepG2 cells with nearly 20% higher than control⁸⁴.

Eighteen membered macrolides

The unique 18-membered macrolide structure with a methyl-substituted ethanoic acid functional side chain strasseriolides A–D 80-83 (**Fig. 5**), were isolated from the endophyte *Strasseria geniculata* CF-247251. The biological evaluation showed potent antiplasmodial activity of the four compounds against *Plasmodium falciparum* 3D7 with IC₅₀ 9.810, 0.013, 0.123, and 0.128 μ M, respectively⁸⁵.



Fig. 5: Sixteen membered macrolides.

Ansa-macrolides

Ansa-macrolides or ansamycins, covered diverse and bioactive natural products that have been isolated mainly from actinomycetes. A characteristic feature of these compounds is the medium to large sized macrolide or macrolactam moiety fused to a mono- or bicyclic aromatic center⁸⁵. The HSP90 inhibitor geldanamycin, is one of the most prominent representatives of this class⁸⁶, in addition to the antimycobacterial antibiotic rifamycin, and the antitumor agent maytansinoid⁸⁷. Ansamacrolides from plant endophytes in the following text and in **Fig. 6**.

The first isolation of ansamacrolides from plant endophytes was the divergolides A-D 84-87 which were isolated from unspecified fungal endophyte from the mangrove *Bruguiera gymnorrhiza*. Divergolide D 87 exhibited strong inhibition of *Mycobacterium vaccae*, while divergolide C 86 was the most active against *B. subtilis* and methicillin resistant *S. aureus*, moreover, it displayed distinct cytotoxicity to lung cancer (LXFA 629L), pancreatic cancer (PANC-1), renal cancer (RXF 486L), and sarcoma (Saos-2)⁸⁸.

Isoindolone-macrolide

They are unusual macrolide skeleton with L-glutamate fragment, an isoindolone unit, and a sesquiterpene moiety. During the last 12 years the only isoindolone macrolides from plant endophytes were the emericellolides A–C 88-90 (**Fig. 6**), which were isolated from the endophytic fungus *Emericella nidulans* HDN12-249⁸⁹.

Isolation and Detection of macrolactone and macrolides

The Mass spectrometry (MS), serving as a universal detection technique, has replaced ultraviolet (UV). fluorometric. and electrochemical detection for multi-macrolide analysis. The chromatographic separation mainly relies on the use of reversed-phase columns. In most studies, a conventional LC with a C_{18} -modified silica stationary phase was used. The mobile-phase composition, concentration, and pH are critical for the optimal ionization and chromatographic separation of macrolides. Acetonitrile and methanol are the mostly used organic solvents in LC or UPLC mobile phases. Either formic acid (0.1%) or ammonium acetate (10-20 mM) often employed as a mobile-phase are modifier⁹⁰. The separation and /or final purification of macrolide molecules usually employ HPLC process, the separation performed on C_{18} reversed phase. The mobile phases A and B are either water and acetonitrile or water and methanol, containing 0.1% formic acid, in a gradient elution (5-100% B for 0-15 min with a linear gradient, followed by 5 min of 100% B) 91,92 .



Fig. 6: Ansa- and Isoindolone-macrolides.

Structure elucidation of macrolides Mass spectrometry

In order to identify and isolate new macrolides from microbial sources, the tandem dereplication is a common mass-based technique for screening of known bioactive compounds. Many databases' platforms are used to reach new macrolides as the Global Natural Products Social Molecular Networking (GNPS), which is an open-access data-driven tandem mass spectral platform, which particularly well-suited and widely used for this purpose. Applying the GNPS Jang et al discovered two new geldanamycin and streptimidone derivatives during their study of Streptomyces species ⁹².

Infrared spectroscopy

The IR spectrum showed absorption for the hydroxy group at 3418 cm⁻¹ and carbonyl carbon at 1732 cm⁻¹, the peaks at 3325 and 1716 cm⁻¹ indicated the presence of hydroxy groups and a carbonyl group. The IR absorption band at 1800 cm⁻¹ identify both α hydroxy- γ -lactone moieties^{72,78}.

Ultraviolet absorption

Due to complicated and variable structure of macrolides, there is no characteristic absorption band or bands to cover the whole class of macrolides, otherwise the absorption varied according to number of chromophores and extent of conjugation in each class of macrolide. Small to medium sized macrolactones (C10-C14), with isolated chromophores have absorption bands at 208-210 nm⁹². β -resorcylic acid derivatives absorption bands are at 211, 262, and 298 nm; 211, 262, 298 nm; 220, 260, 310 or 220, 258, 308⁹¹. Brefeldin A showed absorption band at 230 nm^{24,92}. UV analysis showed a band at 266 nm for Pyrenophorol and two bands at 199, 213 (sh) for tetrahydropyrenophorol. Additionally, Myrothecene analysis revealed the presence of UV absorption bands at 219-220 nm^{71} , meanwhile absorption bands the for Ansamacrolides were at 233, 243 (sh), 254, 282-290 nm^{78,93}.

¹³C NMR spectroscopy

The ¹³C NMR and DEPT spectra have offered a very helpful tool for the major structural features of macrolide with respect to ring size, oxygenated methines, additional carbonyls and sulphur substitution as well as the presence of epoxide moiety 54,92 .

Generally, the ¹³C NMR when combined with HSQC-DEPT data is indicative tool. Surveying the published ¹³C NMR spectral data we could conclude that the resonances ester and/or amide carbonyls at $\delta_{\rm C}$ 170-166, and at $\delta_{\rm C}$ 156.2-153.5, 140-137, 119-115, 110-108, and around 98 for aromatic carbons, in addition to resonances at $\delta_{\rm C}$ 49-34 indicates and identifies the number of methines, at δ_C 32- 21 for methylenes, at $\delta_{\rm C}$ 55.5-56.5 for methoxy groups, and the methyl groups at $\delta_{\rm C}$ 16-23 ppm⁶⁵. The ketone carbonyls at $\delta_{\rm C}$ 209-206. While the methines resonance at $\delta_{\rm C}$ 136.3, 133.4 and 128.0, olefinic methines at $\delta_{\rm C}$ 108-106 and oxygenated methines at $\delta_{\rm C}$ 77.3, 75.4, and 68.5, methylenes at $\delta_{\rm C}$ 48.5, 38.1, 37.5, 28.8, and 27.9, and the methyl groups at $\delta_{\rm C}$ 19- 12^{80} . Meanwhile the carbon resonance at $\delta_{\rm C}$ 61.9 and 63.8 together with the proton at $\delta_{\rm H}$ 4.42 (m) and 4.85 (m) ppm were diagnostic for a 1,2-disubstituted epoxide, while $\delta_{\rm C}$ 41.7 referred to sulphur bearing 43.4 ppm methine^{26,44}.

Conclusion

Since the inspiring isolation of numerous promising plant metabolites as camptothecin and taxol from plant endophytic fungi, they have gained increasing importance as a source of biologically potential active metabolites. Plant endophytes might be the answer to the problem of inadequate medicinal resources and slow growth rates of medicinal plants. According to the current study, research for novel natural products from plant endophytes in the past 10 years had afforded 90 macrolides, including 80 new molecules. The majority of the macrolides described in this study had 12 and 16-membered ring structures, with a few having 10, 13, 14, and 18-membered rings (Fig. 7). They showed cytotoxic, antibacterial, antifungal, and α -glucosidase inhibitory properties (Fig. 8).

The described compounds were reported in 30 fungal strains of 12 genera. Most of the investigated species belonging to the *Cladosporium* and *Penicilium* genera. (Fig.9). It is notable that the number of novel compounds from plant endophytes is much fewer than those reported from marine endophytes during the same time period.



Fig. 7: Ring sizes of the new macrolides isolated during 2013 to 2023.



Fig. 8: Major biological activities of the macrolides isolated during 2013 to 2023.



Fig. 9: Main genera of plant endophytic fungi examined between 2013 to 2023.

We addressed the standard isolation processes of macrolactones and macrolides, as well as a synopsis of the reported compounds' ¹³C-NMR spectroscopic data, as an assistance

in structural elucidation of this class. During the preparation of this review, we observed that the name thiocladospolide F identifies two different structures, as well as the name thiocladospolide G also identifies two different structures, accordingly, we propose that compounds 30 and 31 to be recognized as thiocladospolides F and G, while compounds 35 and 36 should be recognozed as thiocladospolides K and L.

Acknowledgement

This work was supported by National Institutes of Health grant R01 GM115261, the Center of Biomedical Research Excellence (COBRE) in Pharmaceutical Research and Innovation (CPRI, NIH P20 GM130456), the University of Kentucky College of Pharmacy, and the National Center for Advancing Translational Sciences (UL1TR000117 and UL1TR001998).

REFERENCES

- 1. A. Singh, D.K. Singh, R.N. Kharwar, J.F. White and S.K. Gond, "Fungal endophytes as efficient sources of plant-derived bioactive compounds and their prospective applications in natural product drug discovery: Insights, avenues, and challenges", *Microorganisms*, 9(1), 197 (2021).
- H. Gao, G. Li and H.X. Lou, "Structural Diversity and Biological Activities of Novel Secondary Metabolites from Endophytes", *Molecules*, 23(3), 646 (2018).
- M. E. Bungihan, M. A. Tan, H. Takayama, D. Cruz and G. Nonato, "A new macrolide isolated from the endophytic fungus *Colletotrichum* sp.", *Philipp Sci Lett*, 6(1), 57 (2013).
- O.A. Aleynova and K.V. Kiselev, "Interaction of Plants and Endophytic Microorganisms: Molecular Aspects, Biological Functions, Community Composition, and Practical Applications", *Plants*, 12(4), 714 (2023).
- E. S. El-Khayat, R. S. M. Ibrahim and G. A. Mohamed, "Plant endophytes, renewable source of new natural products", *Nat Prod J*, 2(3), 225-234 (2012).
- 6. R.M.K. Toghueo, "Bioprospecting endophytic fungi from *Fusarium* genus as

sources of bioactive metabolites", *Mycology*, 11(1), 1-21 (2020).

- A.E. Fadiji and O.O. Babalola, "Elucidating mechanisms of endophytes used in plant protection and other bioactivities with multifunctional prospects", *Front Bioeng Biotechnol*, 8, 467 (2020).
- Y. Lee, J. Y. Choi, H. Fu, C. Harvey, S. Ravindran, W. R. Roush, J. C. Boothroyd and C. Khosla, "Chemistry and biology of macrolide antiparasitic agents", *J Med Chem*, 54(8), 2792-2804 (2011).
- M.C. Manganyi and C.N. Ateba, "Untapped potentials of endophytic fungi: A review of novel bioactive compounds with biological applications", *Microorganisms*, 8(12), 1934 (2020).
- G. E. S. Batiha, M. A. Zayed, A. A. Awad, H. M. Shaheen, S. Mustapha, O. Herrera-Calderon, J. P. Pagnossa, A. M. Algammal, M. Zahoor and A. Adhikari, "Management of SARS-COV-2 infection: key focus in macrolides efficacy for COVID-19", *Front Med (Lausanne)*, 8, 642313, (2021).
- A. Pani, M.A. Lauriola, F. Romandini and F. Scaglione, "Macrolides and viral infections: focus on azithromycin in COVID-19 pathology", *Int J Antimicrob Agents*, 56(2), 106053 (2020).
- X. Pani, X. Lin, D. Cao, X. Zeng, P.S. Yu, I. He, R. Nussinov and F. Cheng, "Deep learning for drug repurposing: Methods, databases, and applications", *Wiley Interdiscip Rev Comput Mol Sci*, 12(4), e1597 (2022).
- M.A. Islam, M.K. Kibria, M.B. Hossen, M.S. Reza, S.A. Tasmia, K.F. Tuly, M.P. Mosharof, S.R. Kabir, M.H. Kabir and M.N.H. Mollah, "Bioinformatics-based investigation on the genetic influence between SARS-CoV-2 infections and idiopathic pulmonary fibrosis (IPF) diseases, and drug repurposing", *Sci Rep*, 13 (1), 4685 (2023).
- H. Zhang, J. Zou, X. Yan, J. Chen, X. Cao, J. Wu, Y. Liu and T. Wang, "Marinederived macrolides 1990–2020: An overview of chemical and biological diversity", *Mar Drugs*, 19(4), 180 (2021).

- 15. A. Evidente, "Fungal bioactive macrolides", *Nat Prod Rep*, 39(8), 1591-1621 (2022).
- K.D. Lenz, K.E. Klosterman, H. Mukundan and J.Z. Kubicek-Sutherland, "Macrolides: from toxins to therapeutics", *Toxins*, 13(5), 347 (2021).
- G. Dräger, A. Kirschning, R. Thiericke and M. Zerlin, "Decanolides, 10membered lactones of natural origin", *Nat Prod Rep*, 13(5), 365-375 (1996).
- J. Xu, C. Jiang, ZI. Zhang, W.q. Ma and Y.W. Guo, "Recent progress regarding the bioactivities, biosynthesis and synthesis of naturally occurring resorcinolic macrolides.", *Acta Pharm Sin*, 35(3), 316-330 (2014).
- B. Arsic, J. Barber, A. Čikoš, M. Mladenovic, N. Stankovic and P. Novak, "16-membered macrolide antibiotics: a review", *Int J Antimicrob Agents*, 51(3), 283-298 (2018).
- 20. A. Janas and P. Przybylski, "14-and 15membered lactone macrolides and their analogues and hybrids: Structure, molecular mechanism of action and biological activity", *Eur J Med Chem*, 182, 111662 (2019).
- S. Bang, J. Kim, J. Oh, J.S. Kim, S.R. Yu,
 S. Deyrup, Y.S. Bahn and S.H. Shim, "Rare β-Resorcylic Acid Derivatives from a Halophyte-Associated Fungus *Colletotrichum gloeosporioides* JS0419 and Their Antifungal Activities", *Mar Drugs*, 20(3), 195 (2022).
- H. Greve, P.J. Schupp, E. Eguereva, S. Kehraus and G.M. König, "Ten-membered lactones from the marine-derived fungus *Curvularia* sp", *J Nat Prod*, 71(9), 1651-1653 (2008).
- A. Evidente, A. Cimmino, A. Berestetskiy, G. Mitina, A. Andolfi and A. Motta, "Stagonolides B– F, nonenolides produced by Stagonospora cirsii, a potential mycoherbicide of *Cirsium arvense*", *J Nat Prod*, 71(1), 31-34 (2008).
- J. Liang, G. Li, S. Zhou, M. Zhao and L. Cai, "Myrothecium-like new species from turfgrasses and associated rhizosphere", *MycoKeys*, 51, 29-53 (2019).
- 25. S. Ghanty, P.R. Khan and B.S. Reddy, "Stereoselective Total Syntheses of

(3R,5R)-Sonnerlactone and (3R,5S)-Sonnerlactone", *Nat Prod Commun*, 12(9), 1479-1482 (2017).

- V. Rukachaisirikul, S. Pramjit, C. Pakawatchai, M. Isaka and S. Supothina, "10-Membered Macrolides from the Insect Pathogenic Fungus *Cordyceps militaris* BCC 2816", *J Nat Prod*, 67(11), 1953-1955 (2004).
- 27. T.H. Quang, D.C. Kim, P. Van Kiem, C. Van Minh, N.X. Nhiem, B.H. Tai, P.H. Yen, N. Thi Thanh Ngan and H.J. Kim, "Macrolide and phenolic metabolites from the marine-derived fungus Paraconiothyrium sp. VK-13 with anti-inflammatory activity", *J Antibiot*, 71(9), 826-830 (2018).
- 28. K. Ye. H.L. Ai and J.K. Liu. "Identification and bioactivities of secondary metabolites derived from endophytic fungi isolated from ethnomedicinal plants of tujia in hubei province: a review" , Nat Product Bioprospect, 11(2), 185-205 (2021).
- 29. T.N. Clark, A.I. Bishop, M. McLaughlin, L.A. Calhoun, J.A. Johnson and C.A. Gray, "Isolation of (–)–avenaciolide as antifungal and antimycobacterial constituent of a *Seimatosporium* sp. endophyte from the medicinal plant *Hypericum perforatum*", *Nat Prod Commun*, 9(10),1495-1496 (2014).
- N.T. Hiep, Y.H. Choi, N. Kim, S.S. Hong, S.B. Hong, B.Y. Hwang, H.J. Lee, S.J. Lee and D.S. Jang, "Polyhydroxylated macrolides from Seimatosporium discosioides and their effects on activation of peroxisome proliferator-activated receptor gamma", *J Nat Prod*, 75(4), 784-788 (2012).
- H. E. Ortega, Y. Y. Shen, K. TenDyke and L. Cubilla-Ríos, "Polyhydroxylated macrolide isolated from the endophytic fungus *Pestalotiopsis mangiferae*", *Tetrahedron Letters*, 55, 2642 (2014).
- W.B. Botero, M.R. Amorim, I.Z. Carlos, M.C. Polesi and L.C. Santos, "Aromatic Polyketides and Macrolides from *Microsphaeropsis arundinis*", *J Braz Chem Soc*, 31, 364 (2020).
- 33. M. Tsuda, T. Mugishima, K. Komatsu, T. Sone, M. Tanaka, Y. Mikam and J.

Kobayashi, "Modiolides A and B, two new 10-membered macrolides from a marine-derived fungus", *J Nat Prod*, 66(3), 412-415 (2003).

- N.S. Pedra, Kd.C.A. Galdino, D.S. da Silva, P.T. Ramos, N.P. Bona, M.S.P. Soares, J.H. Azambuja, K.M. Canuto, E.Sd. Brito and P.R.V. Ribeiro, "Endophytic fungus isolated from Achyrocline satureioides exhibits selective Antiglioma activity—the role of Sch-642305", *Front Oncol*, 8, 476 (2018).
- 35. W. Ebrahim, A.H. Aly, A. Mándi, F. Totzke, M.H. Kubbutat, V. Wray, W.H. Lin, H. Dai, P. Proksch and T. Kurtán, "Decalactone derivatives from *Corynespora cassiicola*, an endophytic fungus of the mangrove plant *Laguncularia racemosa*", *Eur J Org Chem*, 18, 3476 (2012).
- 36. X.Y. Zhang, Z.L. Liu, B.D. Sun, S.B. Niu, M.H. Wang, X.M. Tan, Z.M. Zou and G. Ding, "Bioactive resorcylic acid lactones with different ring systems from desert plant endophytic fungus *Chaetosphaeronema hispidulur.*", *J Agric Food Chem*, 66(34), 8976-8982 (2018).
- M. Zheng, Z.L. Xu, R.M. Yang, S.C. Hu, G. Ding and Y.G. Zhang, "Stereochemical determination of four 10-membered ring resorcylic acid lactones from the desert plant endophytic fungus *Chaetosphaeronema hispidulum*", J Antibiot, 73(7), 471-474 (2020).
- 38. A. Sugawara, A. Sueki, T. Hirose, H. Shima, K.S. Akagawa, S. Ōmura and T. Sunazuka, "Novel 12-membered non-antibiotic macrolides, EM900 series with anti-inflammatory and/or immunomodulatory activity; synthesis, structure–activity relationships and in vivo study", *J Antibiot*, 65(9), 487-490 (2012).
- 39. F.Z. Zhang, X.M. Li, X. Li, S.Q.Yang, L.H. Meng and B.G. Wang, "Polyketides from the mangrove-derived endophytic fungus *Cladosporium cladosporioides*", *Mar Drugs*, 17(5), 296 (2019).
- C.J. Smith, D. Abbanat, V.S. Bernan, W.M. Maiese, J. Jompa, A. Tahir and C.M. Ireland, "Novel polyketide metabolites from a species of marine fungi", *J Nat Prod*, 63(1), 142-145(2000).

- 41. H.X. Liu, H.B. Tan, S.N. Li, Y.C. Chen, H.H. Li, S.X. Qiu and W.M. Zhang, "Two new 12-membered macrolides from the endophytic fungal strain *Cladosprium colocasiae* A801 of *Callistemon viminalis*", *J Asian Nat Prod Res*, 21(7), 696-701(2019).
- Wuringege, Z.K. Guo, W. Wei, R.H. Jiao, T. Yan, L.Y. Zang, R. Jiang, R.X. Tan and H.M. Ge, "Polyketides from the plant endophytic fungus *Cladosporium* sp. IFB3lp-2", *J Asian Nat Prod Res*, 15(9), 928-933 (2013).
- F.Z. Zhang, X.M. Li, S.Q. Yang, L.H. Meng and B.G. Wang, "Thiocladospolides A–D, 12-membered macrolides from the mangrove-derived endophytic fungus *Cladosporium cladosporioides* MA-299 and structure revision of pandangolide 3", *J Na Prod*, 82(6), 1535-1541 (2019).
- 44. W. Wang, H. Feng, C. Sun, Q. Che, G. Zhang, T. Zhu and D. Li, "Thiocladospolides FJ, antibacterial sulfur containing 12-membered macrolides from the mangrove endophytic fungus *Cladosporium oxysporum* HDN13-314", *Phytochemistry*, 178, 112462 (2020).
- 45. Y.H. Wu, Z.H. Zhang, Y. Zhong, J.J. Huang, X.X. Li, J.Y. Jiang, Y.Y. Deng, L.H. Zhang and F. He, "Sumalactones A– D, four new curvularin-type macrolides from a marine deep sea fungus *Penicillium Sumatrense*", *RSC Adv*, 7(63), 40015-40019 (2017).
- 46. D.J. Robeson, G.A. Strobel and R.N. Strange, "The identification of a major phytotoxic component from *Alternaria macrospora* as α,β -dehydrocurvularin", *J Nat Prod*, 48(1), 139–141 (1985).
- 47. B.P. Bashyal, E.K. Wijeratne, J. Tillotson, A.E. Arnold, E. Chapman and A.L. Gunatilaka, "Chlorinated dehydrocurvularins and alterperylenepoxide A from *Alternaria* sp. AST0039, a fungal endophyte of *Astragalus lentiginosus*", J Nat Prod, 80(2), 427-433(2017).
- 48. CJ Barrow, "New macrocyclic lactones from a *Penicillium* species", *J Nat Prod*, 60(10), 1023–1025 (1997).
- 49. L.H. Meng, X.M. Li, C.T. Lv, C.S. Li, G.M. Xu, C.G. Huang and B.G. Wang,

"Sulfur-containing cytotoxic curvularin macrolides from Penicillium sumatrense MA-92, a fungus obtained from the rhizosphere of the mangrove *Lumnitzera racemosa*", *J Nat Prod*, 76(11), 2145–2149 (2013).

- 50. F.Z. Zhang, X.M. Li, L.H. Meng and B.G. Wang, "Cladocladosin A, an unusual macrolide with bicyclo 5/9 ring system, and two thiomacrolides from the marine mangrove-derived endophytic fungus, *Cladosporium cladosporioides* MA-299", *Bioorg Chem*, 101, 103950 (2020).
- 51. N. Jana and S. Nanda, "Resorcylic acid lactones (RALs) and their structural congeners: recent advances in their biosynthesis, chemical synthesis and biology", *New J Chem*, 42(22), 17803-17873 (2018).
- 52. L. Xu, Z. He, J. Xue, X. Chen and X. Wei, "β-Resorcylic acid lactones from a Paecilomyces fungus", *J Nat Prod*, 73(5), 885–889 (2010).
- 53. M. Buayairaksa, S. Kanokmedhakul, K. Kanokmedhakul, P. Moosophon, C. Hahnvajanawong and K. Soytong, "Cytotoxic lasiodiplodin derivatives from the fungus Syncephalastrum racemosum", *Arch Pharm Res*, 34(12), 2037-2041 (2011).
- 54. S. Chen, Z. Liu, H. Li, G. Xia, Y. Lu, L. He, S. Huang and Z. She, "β-Resorcylic acid derivatives with α-glucosidase inhibitory activity from *Lasiodiplodia* sp. ZJ-HQ1, an endophytic fungus in the medicinal plant Acanthus ilicifolius", *Phytochem Lett*, 13, 141-146 (2015).
- 55. A.A. Stierle, D.B. Stierle, D. Decato, N.D. Priestley, J.B. Alverson, J. Hoody, K. McGrath and D. Klepacki, "The berkeleylactones, antibiotic macrolides from fungal coculture", *J Nat Prod*, 80(4), 1150 (2017).
- 56. B. Ferko, M. Zeman, M. Formica, Sn Veselý, J. Doháňošová, P. Olejníková, Da Berkeš, P. Jakubec and D.J. Dixon, "Total synthesis of berkeleylactone A", *J Org Chem*, 84(11), 7159–7165 (2019).
- N.O. Anadu, V.J. Davisson and M. Cushman, "Synthesis and anticancer activity of brefeldin A ester derivatives", *J Med Chem*, 49(13), 3897-3905 (2006).

- 58. J. Xie, Y.Y. Wu, T.Y. Zhang, M.Y. Zhang, W.W. Zhu, E.A. Gullen, Z.J. Wang, Y.C. Cheng and Y.X. Zhang, "New and bioactive natural products from an endophyte of *Panax notoginseng*", *RSC Adv*, 7(60), 38100 (2017).
- 59. N. Gao, Z.C. Shang, P. Yu, J. Luo, K.L. Jian, L.Y. Kong and M.H. Yang, "Alkaloids from the endophytic fungus *Penicillium brefeldianum* and their cytotoxic activities", *Chin Chem Lett*, 28(6), 1194-1199 (2017).
- 60 A. Ballio, A. Evidente, A. Graniti and L. Sparapano, "Seiricuprolide, a new phytotoxic macrolide from a strain of Seiridium cupressi infecting cypress", *Phytochemistry*, 27(10), 3117-3121 (1988).
- 61. A. Zinedine, J.M. Soriano, J.C. Molto and J. Manes, "Review on the toxicity, occurance, metabolism, detoxification, regulations and intake of Zearalenone: an oestrogenic mycotoxin", *Food Chem Toxicology*, 45(1), 1-18 (2007).
- 62. K. Kito, R. Ookura, S. Yoshida, M. Namikoshi, T. Ooi and T. Kusumi, "New cytotoxic 14-membered macrolides from marine-derived fungus Aspergillus ostianus", *Org Lett*, 10(2), 225-228 (2008).
- H.X. Liu, W.Z. Liu, Y.C. Chen, Z.H. Sun, Y.Z. Tan, H.H. Li and W.M. Zhang, "Cytotoxic trichothecene macrolides from the endophyte fungus Myrothecium roridum", *J Asian Nat Prod Res*, 18(7), 684-689 (2016).
- 64. J.S. Harwooda, H.G. Cutler and J.M. Jacyno, "Nigrosporolide, a plant growthinhibiting macrolide from the mould *Nigrospora sphaerica*", *Nat Prod Lett*, 6(3), 181 (1995).
- S. Liu, H. Dai, G. Makhloufi, C. Heering, C. Janiak, R. Hartmann, A. Mandi, T. Kurtán, W. E. Müller, and M. U. Kassack, "Cytotoxic 14-membered macrolides from a mangrove-derived endophytic fungus, Pestalotiopsis microspora", *J Nat Prod*, 79(9), 2332-2340 (2016).
- 66. R.N. Asolkar, R.P. Maskey, E. Helmke, H. Laatsch, Chalcomycin B, a new macrolide antibiotic from the marine isolate

Streptomyces sp. B7064., *J. Antibiot.*, 55(10), 893 (2002).

- 67. S. Carlson, L. Marler, S.J. Nam, B.D. Santarsiero, J.M. Pezzuto and B.T. Murphy, "Potential chemopreventive activity of a new macrolide antibiotic from a marine-derived *Micromonospora* sp", *Mar Drugs*, 11(4), 1152-1161 (2013).
- 68. P. Zarogoulidis, N. Papanas, I. Kioumis, E. Chatzaki, E. Maltezos and Κ. Zarogoulidis, "Macrolides: from in vitro anti-inflammatory and immunomodulatory properties to clinical practice in respiratory diseases", Eur J Clin Pharmacol, 68(5), 479-503 (2012).
- M.W. Sumarah, J.R. Kesting, D. Sørensen and J.D. Miller, "Antifungal metabolites from fungal endophytes of Pinus strobus", *Phytochemistry*, 72(14-15),1833-1837 (2011).
- M.W. Sumarah, A.K. Walker, K.A. Seifert, A. Todorov, J.D. Miller, "Screening of fungal endophytes isolated from eastern white pine needles", in: Jetter R (ed). The Formation, Structure and Activity of Phytochemicals. Springer, Switzerland, 195 -206 (2015).
- 71. K. Krohn, U. Farooq, U. Flörke, B. Schulz, S. Draeger, G. Pescitelli, S. Antus and T. Kurtán, "Secondary Metabolites Isolated from an Endophytic *Phoma* sp. Absolute Configuration of Tetrahydropyrenophorol Using the Solid-State TDDFT CD Methodology", *Eur J Org Chem*, 19, 3206-3211 (2007).
- 72. L. Shen, J.S. Wang, H.J. Shen, Y.C. Song and R.X. Tan, "A new cytotoxic trichothecene macrolide from the endophyte *Myrothecium roridum*", *Planta Medica*, 76(10), 1004-1006 (2010).
- 73. J. He, T. Zhou, J.C. Young, G.J. Boland, P.M. Scott, Chemical and biological transformations for detoxification of trichothecene mycotoxins in human and animal food chains: a review, *Trends Food Sci Tech*, 21(2), 67 (2010).
- 74. T. Amagata, C. Rath, J.F. Rigot, N. Tarlov, K. Tenney, F.A. Valeriote and P. Crews, "Structures and Cytotoxic Properties of Trichoverroids and Their Macrolide Analogues Produced by Saltwater Culture of Myrothecium

verrucaria", *J Med Chem*, 46(20), 4342-4350 (2003).

- 75. G.S. Bondy and J.J. Pestka, "Immunomodulation by fungal toxins", J *Toxicol Environ Health B Crit Rev*, 3(2), 109-143 (2000).
- 76. H. Abbas, B. Johnson, W. Shier, H. Tak, B. Jarvis and C. Boyette, "Phytotoxicity and mammalian cytotoxicity of macrocyclic trichothecene mycotoxins from Myrothecium verrucaria", *Phytochemistry*, 59(3), 309-313 (2002).
- T. Sugawara, A. Tanaka, K. Nagai, K. Suzuki and G. Okada, "New members of the trichothecene family", *J Antibiot*, 50(9), 778-780 (1997).
- M. Isaka, J. Punya, Y. Lertwerawat and Y. Thebtaranonth, "Antimalarial activity of macrocyclic trichothecenes isolated from the fungus *myrothecium verrucaria*", *J Nat Prod*, 62(2), 329-331 (1999).
- 79. L. Shen, C.Z. Ai, Y.C. Song, F.W. Wang, R.H. Jiao, A.-H. Zhang, H.Z. Man and R.X. Tan, "Cytotoxic trichothecene macrolides produced by the endophytic *Myrothecium roridum*", *J Nat Prod*, 82(6), 1503-1509 (2019).
- H. Liu, Y. Chen, S. Li, W. Zhang, Z. Liu and W. Zhang, "Trichothecene macrolides from the endophytic fungus Paramyrothecium roridum and their cytotoxic activity", *Fitoterapia*, 147, 104768 (2020).
- L. Shen, L. Zhu, Q. Tan, D. Wan, J. Xie and J. Peng, "New cytotoxic trichothecene macrolide epimers from endophytic *Myrothecium roridum* IFB-E012", J *Antibiot*, 69(8), 652-655 (2016).
- A. Shimada, S. Takeuchi, M. Kusano, S. Fujioka and Y. Kimura, "Roridin A, Verrucarin A, inhibitors of pollen development in Arabidopsis thaliana, produced by *Cylindrocarpon* sp", *Plant Sci J*, 166(5), 1307-1312 (2004).
- 83. L.T.T. Nguyen, J.Y. Jang, T.Y. Kim, N.H. Yu, A.R. Park, S. Lee, C. H. Bae, J.H. Yeo, J.S. Hur and H.W. Park, "Nematicidal activity of verrucarin A and roridin A isolated from *Myrothecium verrucaria* against Meloidogyne incognita", *Pestic Biochem Physiol*, 148, 133-143 (2018).

- 84. T. Lin, G. Wang, Y. Zhou, D. Zeng, X. Liu, R. Ding, X. Jiang, D. Zhu, W. Shan and H. Chen, "Structure elucidation and biological activity of two new trichothecenes from an endophyte, *Myrothecium roridum*", J Agric Food Chem, 62(25), 5993-6000 (2014).
- 85. F. Annang, G. Pérez-Moreno, V. González-Menéndez, R. Lacret, I. Pérez-Victoria, J. Martín, J. Cantizani, N. de Pedro, D. Choquesillo-Lazarte and L.M. Ruiz-Pérez, "Strasseriolides A–D, a family of antiplasmodial macrolides isolated from the fungus *Strasseria geniculata* CF-247251", *Org Lett*, 22(17), 6709–6713 (2020).
- 86. L. Ding, A. Maier, H.H. Fiebig, H. Görls, W.H. Lin, G. Peschel and C. Hertweck, "Divergolides A–D from a Mangrove Endophyte Reveal an Unparalleled Plasticity in ansa-Macrolide Biosynthesis", *Angew Chem*, 123(7), 1668 (2011).
- Y. Fukuyo, C.R. Hunt and N. Horikoshi, "Geldanamycin and its anti-cancer activities", *Cancer Lett*, 290(1), 35-24 (2010).
- L. Ding, A. Maier, H.H. Fiebig, W.H. Lin and C. Hertweck, "A family of multicyclic indolosesquiterpenes from a bacterial endophyte", *Org Biomol Chem*, 9(11), 4029 (2011).

- H. Zhou, X. Sun, N. Li, Q. Che, T. Zhu, Q. Gu and D. Li, "Isoindolone-containing meroperpenoids from the endophytic fungus *Emericella nidulans* HDN12-249", *Org Lett*, 18(18),4670-4573 (2016).
- 90. J. Wang, "Analysis of Macrolide Antibiotics, Using Liquid Chromatography-Mass Spectrometry, in Food, Biological and Environmental Matrices", *Mass Spectrom Rev*, 28(1),50-92. (2009).
- I. Kanfer, M. F. Skinner and R. B. Walker, "Analysis of Macrolides Antibiotic", J Chromatogr A, 812(1-2),255-286 (1998).
- 92. S. Bang and S.H. Shim, "Beta resorcylic acid lactones (RALs) from fungi: chemistry, biology, and biosynthesis", *Arch Pharm Res*, 43(11), 1093-1113 (2020).
- 93. S. M. Kupchan, Y. Komoda, W. Court, G. Thomas, R. Smith, A. Karim, C. Gilmore, R. Haltiwanger and R. Bryan, "Maytansine, a novel antileukemic ansa macrolide from Maytenus ovatus", *J Am Chem Soc*, 94(4), 1354-1356 (1972).

Bull. Pharm. Sci., Assiut University, Vol. 47, Issue 1, 2024, pp. 151-168

