PHYTOCHEMICAL STUDY OF *IXORA FINLAYSONIANA* WALL. EX.G.DON GROWING IN EGYPT

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نبات إيكسورا فينلانسونيانا وال.إكس جى.دون التابع للعائلة الفوية هو نبات زينة دائم الخضرة معروف بجمال أز هاره وله العديد من الاستعمالات الشعبية و الطبية. و تمت دراسة المحتويات الكيميائية لهذا النبات وتم فصل ثلاثة مركبات من خلاصة الهكسان وهم: ن-نوناكوسانول، ألفا أميرين و بيتاسيتوستيرول. ومن خلاصة الكلوروفورم تم فصل أربعةمركبات وهى: 3-هيدروكسى هكسان-5-أوليد، حمض بروتوكاتيشيك، حمض الجاليك و 5-أبيتا-سيتوستيرول جليكوزيد بالإضافة إلى ثلاثة مركبات أخرى من خلاصة خلات الإيثيل وهم: بار اسور بوسيد، دى-وجالاكتيتول. وقد تم التعرف على تركيب هذه المركبات بدراسة الخواص الطبيعية و الكيميائية و والطيفية المختلفة لكل منها.

A hydrocarbon alcohol, nonacosanol (1), α -amyrin (2) and β -sitosterol (3) were isolated from the n-hexane fraction of the methanolic extract of Ixora finlaysoniana. From the chloroform fraction, four compounds were isolated; 3-hydroxyhexan-5-olide (4), protocatechuic acid (5), gallic acid (6) and β -sitostreol glucoside (7). In addition to three compounds were isolated form the ethyl acetate fraction; parasorboside (8), D-1-O-methylmyo-inostiol (9) and galactitol (10). The structures of the isolated compounds were elucidated on the basis of physical, chemical and spectroscopic evidence.

INTRODUCTION

The genus Ixora (Rubiaceae) comprises about 200 species distributed through the warm regions in the tropics.¹⁻³ Ixora finlaysoniana Wall.ex.G.Don. is a handsome woody shrub with showy flowers in clusters and evergreen foliage.^{3,4} Though, the plants of the genus *Ixora* are cultivated for ornamental purposes, some species have been used in traditional medicine. Ixora wood is febrifuge and the root is aperient, diuretic and deobstruent.⁵ The flowers of I. coccinea L. have been used against leucorrhea and catarrhal bronchitis⁶ and for dysentery, healing of ulcers and have antiinflammatory and antitumor activities.7,8 The plant also showed chemoprotective effect on cyclophosphamide-induced toxicity by increasing the life span of treated mice.9 I. chinensis is used to treat headache and stomachache and as a remedy for incipient tuberculosis.⁶ The dried entire plant of Ixora finlaysoniana is used in Thailand as a strength

medicine,¹⁰ while the ethanolic extract of the plant was proved to have estrogenic, abortifacient and anti-implantation effects.¹¹

The chemical composition of some *Ixora* species have been carried out. Phenolic acids and flavonoidal glycosides were isolated from *I. Undulata*,¹² while chrysin-5-*O*- β -D-xylopyranoside was isolated from the stem of *I. arborea*.¹³ A C₁₈ conjugated tetraenoic acid was isolated from *I. chinensis* seed oil.¹⁴ Additionally, some antitumor principles, mainly phenolic acids, were isolated from *I. Javanica*.¹⁵

Reviewing the current literature for the chemical composition of *Ixora finlaysoniana*¹⁶ recovered only the isolation of apigenin-4'-O- β -D-glucopyranoside and 11-hydroxy-dodec-5-en-2-one. The botanical study of the stem and leaf of *Ixora finlaysoniana* cultivated in Egypt was previously carried out.¹⁷

EXPERIMENTAL

I- Plant material

The plant material used in this work was collected in Aswan Botanical Garden, Aswan, Egypt in March 1998. The plant was kindly authenticated by the Royal Botanical Garden KEW. A voucher specimen was deposited in the Herbarium of the Pharmacognosy Department, Faculty of Pharmacy, Assiut University, Assiut.

II- General procedure

- 1- Melting points were determined using electrothermal digital instrument without correction.
- 2- EIMS were recorded on a JEOL mass. spectrometer at 70 ev.
- 3- ¹H and ¹³C-NMR were recorded on a JEOL spectrometer at 400 and 270 MHz for ¹H-NMR and 100 and 67.5 MHz for ¹³C-NMR using DMSO and CD₃OD as solvent, chemical shift values are reported in δ ppm relative to that of TMS as internal standard and coupling constants are given in Hz.
- 4- Column chromatography was carried out on silica gel 60 (E-Merck, 70-230 mesh) and prepacked column for MPLC (CIG column system, 22 mm, i.d. x 30 cm, Kusano Scientific Co. Tokyo, Japan). Monitoring with TLC was carried out on precoated sheets (Kieselgel 60 F_{254} , Merck). The spots were visualized by spraying with 10% aqueous sulfuric acid and heating at 110°.
- 5- Solvent systems
 - 1- CHCl₃-C₆H₆ (5:1)
 - 2- CHCl₃-MeOH (12:1)
 - 3- CHCl₃-MeOH (8:1)
 - 4- CHCl₃-MeOH (5:1)
 - 5- CHCl₃-MeOH-H₂O (3:1:0.1)

III- Extraction and isolation

The air-dried leaves and stems of *Ixora finlaysoniana* (600 g) were extracted by percolation with methanol for three times (4 L, each) at room temperature. The combined methanol extracts were evaporated under *vacuum* to yield crude extract (60 g). The dried methanol extract was suspended in distilled water (1 L) and partitioned with n-C₆H₁₂ (1L x 3), CHCl₃ (1Lx3) and EtOAc (1L x 3), successively. Each extract was evaporated under *vacuum* till dryness.

The n-C₆H₁₂ fraction (10 g, dark green residue) was subjected to silica gel column chromatography (400 g) using gradient n-C₆H₁₂-CHCl₃ system. The fractions 22-27 (1 g, n-C₆H₁₂-CHCl₃ 65:35) were further purified by repeated silica gel column chromatography to give compounds **1** (50 mg, R_f 0.83, solvent system 1), **2** (70 mg, R_f 0.80, solvent system 2) and **3** (300 mg R_f 0.66, solvent system2).

The CHCl₃ fraction (10 g. faint brown residue) was subjected to silica gel column chromatography (400 g) using gradient CHCl₃-MeOH system. The fractions 20-24 (500 mg, CHCl₃-MeOH 85:15) were subjected to repeated moderate pressure column chromatography (MPLC), [silica gel prepacked column] using CHCl₃-MeOH (9:1) to give compounds 4 (100 mg, Rf 0.52, solvent system 2) and 5 (120 mg, R_f 0.61, solvent system 3). Fractions 32-40 (2 g, CHCl₃-MeOH 8:2) were further purified by repeated silica gel column chromatography using gradient CHCl₃-MeOH to give compound 6 (500 mg, $R_f 0.54$, solvent system 3) and compound 7 (350 mg, R_f 0.50, solvent system 3).

The EtOAc fraction (15 g, yellowish brown residue) was chromatographed over silica gel column (500 g), elution was carried out with CHCl₃ increasing the polarity with MeOH gradiently up to (1:1). Fractions 14-19 eluted with CHCl₃-MeOH (85:15) were combined and evaporated under vacuum to give residue (800 mg) which was а rechromatographed over silica gel column with gradient CHCl₃-MeOH system to give compounds 8 (330 mg, R_f: 0.72, solvent system 4) and 9 (100 mg, R_f : 0.67, solvent system 4). Fractions 22-26 eluted with CHCl₃-MeOH (75:25) were combined and evaporated under *vacuum* to give a residue (150 mg) which was rechromatographed over silica gel column with a mixture of CHCl₃-MeOH (3:1) to give compound 10.

n-Nonacosanol (1): White wax, EI-MS, m/z (% rel. int.) 424 $[M]^+$ (3.1) calculated for C₂₉H₆₀O, 385 (3), 365 (4), 337 (6), 314 (7), 284 (8.), 255 (13), 238 (11), 196 (12), 189 (12), 165 (12), 125 (18), 109 (30), 95 (41), 83 (44), 71 (61), 57 (89) and 43 (100). ¹H-NMR (CDCl₃) $\delta_{\rm H}$: 0.87 (t, *J*= 7.2 Hz), 1.25 (m, CH₂), 1.65 (m, CH₂), 2.3 (t, *J*= 7.8 Hz), 4.05 (t, *J*= 6.4 Hz), 4.56 (br. s.). ¹³C-NMR (CDCl₃) $\delta_{\rm C}$: 62.4



Compounds isolated from Ixora finlaysoniana

(t, <u>CH</u>₂OH), 34.4 (t), 31.9-29.7 (t), 22.7 (t, <u>CH</u>₂-CH₃) and 14.1 (q, CH₃).

3-Hydroxyhexan-5-olide (**4**): Faint yellow oil, EIMS, m/z (% rel. int.): 131 $[M+1]^+$ (12), 130 $[(M)^+$, $C_6H_{10}O_3]$ (8), 115 $[M-CH_3]^+$ (5), 113 $[M-OH]^+$ (20), 102 $[M-CO]^+$ (4), 97 (35), 86 $[M-CH_3CHO]^+$ (30), 96 $[M-(CH_3CHO+HO)]^+$ (50), 57 (80), 50 (54) and 43 (100). ¹H-NMR (DMSO) δ_{H} : 1.26 (3H, d, *J*= 6.4 Hz, CH₃-6), 1.58 (1H, ddd, *J*= 14.5, 11.0, 3.0 Hz, H-4 β), 2.30 (1H, m, H-4 α), 2.65 (1H, dd, *J*= 17.2, 4.0 Hz, H-2 β), 2.94 (1H, m, H-2 α) 4.36 (1H, m, H-3) and 4.88 (1H, m, H-5). ¹³C-NMR (DMSO) δ_C : 169.0 (s, C-1), 72.8 (d, C-5), 67.4 (d, C-3), 38.1 (t, C-2), 35.6 (t, C-4) and 20.8 (q, C-6).

Protocatechuic acid (**5**): Faint brown amorphous powder, ¹H-NMR (DMSO), $\delta_{\rm H}$: 12.20 (COOH), 9.20 (OH-3), 9.18 (OH-4), 6.92 (m, H-2, H-5, H-6). ¹³C-NMR (DMSO), $\delta_{\rm C}$: 167.4 (C-7), 145.6 (C-4), 145.1 (C-3), 120.4 (C-1), 110.6 (C-6), 109.3 (C-5) and 108.5 (C-2).

Gallic acid (6): Faint brown crystals, m.p 239-241°, ¹H-NMR (DMSO), $\delta_{\rm H}$: 12.2 (COOH), 9.22 (OH-3, OH-5), 8.94 (OH-4), 6.93 (2H, s, H-2, H-6). ¹³C-NMR (DMSO), $\delta_{\rm C}$: 167.4 (C-7), 145.3 (C-3, C-5), 137.9 (C-4), 120.3 (C-1), 108.0 (C-2, C-6).

Parasorboside (8): White amorphous powder, EIMS, m/z (% rel. int.): 292 $[M]^+$ (4), 131 [M+1 of aglycone]⁺ (70), 130 [(M-162), M-hexose]⁺ (4), 115 (15), 113 (5), 102 (90), 97 (19), 86 (26), 73 (100), 69 (56), 57 (87), 55 (68) and 43 (97). ¹H-NMR (DMSO), $\delta_{\rm H}$: 1.26 (3H, d, J= 6.4 Hz, CH₃-6), 1.61 (1H, ddd, J= 14.2, 11.2, 2.8 Hz, H-4β), 2.26 (1H, m, H-4α), 2.89 (1H, dd, J= 17.5, 4.0 Hz, H-2 β), 3.04 (1H, m, H-2α), 4.34 (1H, m, H-3), 4.86 (1H, m, H-5), 5.05 (1H, d, J= 8Hz, H-1' glucose) and 3.75-4.86 (overlapping signals, H-2'-H-6' glucose). ¹³C-NMR (DMSO), δ_{C} : 168.5 (s, C-1), 74.9 (d, C-5), 72.1(d, C-3), 36.5 (t, C-2), 34.2 (t, C-4), 22.4 (q, C-6), 104.0 (d, C-1'), 77.8 (d, C-5'), 77.0 (d, C-3'), 74.3 (d, C-2'), 71.6 (d, C-4'), 62.1 (t, C-6').

D-1-*O*-methyl-*myo*-inositol (**9**): White amorphous powder. EIMS, m/z (% rel. int.) 194 $[M]^+$ (26),179 (72), 163 (11), 162 (12), 145 (10), 102 (31), 98 (42), 85 (75), 54 (100). ¹H-NMR (DMSO) δ_{H} : 3.57 (3H, s, OCH₃), 3.33-3.99 (6H, overlapping signals, H-1 –H-6). ¹³C-NMR (DMSO) δ_{C} : 81.6 (d, C-1), 75.3 (d, C-5), 72.5 (d, C-3), 71.8 (d, C-6), 71.6 (d, C-4), 68.3 (d, C-2), 56.6 (q, OCH₃).

Galactitol (**10**): Colorless prisms, m.p182-185°. EI-MS, m/z (% rel. int.) 182 $[M]^+$ (24), 164 (15), 146 (31), 145 (70), 133 (52), 132 (98), 121 (25),116 (54), 104 (42), 103 (71), 91 (51), 87 (35), 84 (35), 75 (35), 73 (83), 69 (42) and 31 (100). ¹H-NMR (DMSO), δ_{H} : 4.32 (2H, dd, *J*= 11.7, 3.0 Hz, H-1_A, H-6_A), 4.12 (2H, d, *J*= 7.1 Hz, H-3, H-4), 3.60 (2H, m, H-2, H-5), 3.50 (2H, dd, *J* = 11.7, 6.0 Hz, H-1_B, H-6_B). ¹³C-NMR (DMSO), δ_{C} : 71.5 (d, C-3, C-4), 69.9 (d, C-2, C-5), 64.1 (t, C-1, C-6).

RESULTS AND DISCUSSION

Fractionation of the n-hexane fraction of *Ixora finlaysoniana* by repeated column chromatography using silica gel (Experimental section) yielded compounds **1-3**.

Compound 1 showed a molecular ion peak at m/z 424 calculated for $C_{29}H_{60}O$ and was identified from its mass fragmentation pattern, ¹H- and ¹³C-NMR as a straight chain alkanol; n-nonacosanol.¹⁸

Compound 2 was identified as α -amyrin and compound 3 was identified as β -sitosterol through their physical and chromatographic characteristics and by comparison with authentic samples.

The chloroform fraction of the plant was fractionated using a combination of silica gel column chromatography and MPLC (see experimental section) to yield four pure compounds (**4-7**).

The EI-mass spectrum of compound **4** showed $[M+1]^+$ peak at m/z 131 consistent with the molecular formula $C_6H_{10}O_3$, which was, confirmed with ¹³C-NMR and DEPT experiments. The ¹H-NMR spectra exhibited signals for a doublet methyl at δ_H 1.26 (3 H, d, J= 6.4 Hz), two methylene protons at δ_H 1.58 (1H, ddd, J= 14.5, 11, 3.0 Hz) and 2.30 (1H, m), other two methylene protons at δ_H 2.65 (1H, dd, J= 17.2,4.0 Hz) and 2.94 (1H, m) and two oxygen bonded methine protons at δ_H 4.36

(1H, m) and 4.88 (1H, m). Furthermore, the ¹³C-NMR spectra showed signal for a methyl group at $\delta_{\rm C}$ 20.8 (q),two signals for methylene carbons at δ_C 38.1(t) and 35.6 (t), two signals for oxygenated methine carbons at $\delta_{\rm C}$ 72.8 (d) and 67.4 (d) and a lactone carbonyl carbon at $\delta_{\rm C}$ 169.0 (s). Based on these abovementioned data, compound 4 seemed to be a hexanolide¹⁹ with an oxygen substituent and a methyl group which was supported by comparison of its physical and spectral data with those published for other hexanolides.^{19,20} The ¹H-NMR spectra indicated that H-3 appeared at $\delta_{\rm H}$ 4.36 and H-5 at $\delta_{\rm H}$ 4.88 clearly indicated the axial orientation of the former and equatorial orientation of the latter²⁰ and this confirmed the structure of $\mathbf{4}$ as 3-hydroxyhexan-5-olide. This compound was previously isolated from Osmunda japonica^{19,20} but this is the first report for its isolation from the genus Ixora.

Compounds **5** and **6** were identified as protocatechuic acid and gallic acid respectively by comparing their physical and spectral data with those reported, ^{21,22} while compound **7** was identified as sitosterol-3-O- β -D-glucoside by comparing with authentic sample.

The ethyl acetate fraction was fractionated using repeated silica gel column chromatography (see experimental section) to yield three pure compounds (8-10).

The ¹H- and ¹³C-NMR spectra of **8** showed signals closely similar to those of **4** in addition to signals attributed to one hexose moiety. The assignment of the ¹H-NMR and ¹³C-NMR spectral data of **8** is described in the experimental section.

The significant downfield shift of C-3 in ¹³C-NMR of **8** (+ 4.7 ppm) compared with that of **4** together with the upfield shift of C-2 (-1.6 ppm) and C-4 (-1.4 ppm) confirmed the attachment of the hexose moiety to C-3 of **4**.²³ The carbon signals at $\delta_{\rm C}$ 104.0 (d), 77.8 (d), 77.0 (d), 74.3 (3), 71.6(d) and 62.1 (t) and the coupling constant of the anomeric proton in the ¹H-NMR spectra (*J*= 8.0 Hz) proved that the hexose moiety is β -D-glucopyranose.^{23,24}

On the basis of these evidences the structure of **8** was presumed to be (3R, 5R)-3- $(\beta$ -glucopyranosyloxy)-5-hexanolide

(parasorboside) which has been isolated from *Sorbus aucuparia* L.,²⁵ *Osmunda japonica*²⁰ and *Hymenophyllum barbatum*.²⁶ This compound was reported to have a significant

anti-feeding activity for larva of yellow butterfly *Eurema hecabe mandarina*.²⁵

Compound **9** was obtained as an amorphous powder. Its molecular formula was suggested to be $C_7H_{14}O_6$ from its EI-MS 13 C-NMR and DEPT spectra. Its EI-MS showed a molecular ion peak at m/z 194, a base peak at m/z 41 and other major fragment ions at m/z 179, 163, 162, 145, 102, 98, 85 and 54. The ¹H-NMR spectrum of **9** showed a singlet signal at δ_H 3.57 (3H, s) assigned for a methoxy protons in addition to a complex pattern of overlapping signals at δ_H 3.33 – 3.99 equivalent to six methine protons.

The ¹³C-NMR spectrum of **9** indicated the presence of a methoxy group ($\delta_{\rm C}$ 56.6) and six oxymethines [$\delta_{\rm C}$ 68.30 (C-2), 71.75 (C-4), 71.81 (C-6), 72.50 (C-3), 75.30 (C-5) and 81.60 (C-1)]. Comparing the above mentioned data with those reported indicated that these data are characteristic to *myo*-inositol derivatives,^{27,28} since in other inositol derivatives as *scyllol*-inositol, all the carbon signals are identical (appeared at $\delta_{\rm C}$ 73.7)^{23,28} and in L- *chiro*-inositol, only three signals were appeared (at $\delta_{\rm C}$ 70.5, 71.6 and 72.8), while in *epi*-inositol, four signals were appeared (at $\delta_{\rm C}$ 71.7, 74.5, 70.1 and 66.8).^{23,28}

The downfield shift of C-1 of compound **9** ($\delta_{\rm C}$ 81.6) and the upfield shift of C-2 and C-6 together with the appearance of the methoxy group signal at $\delta_{\rm C}$ 56.6 indicated that **9** is D-1-*O*-methyl-*myo*-inositol^{23,28} previously isolated from *Crotalaria trifoliastrum*²⁸ but it is the first time to be isolated from the genus *Ixora*.

Compound 10 was obtained as colorless prisms and exhibited a molecular formula $C_6H_{14}O_6$ as determined by EI-MS and DEPT 13 C-NMR spectra. In the 13 C-NMR and DEPT spectra, only three carbon signals were appeared at δ_C 71.5 (d x 2), 69.9 (d x 2) and 64.1 (t x 2) suggested a hexose moiety.²³ This was confirmed by the EI-MS which showed a molecular ion peak at m/z 182, other peaks at m/z 164 and 146 for successive loss of water molecules and a base peak at m/z 31 for CH₂OH⁺ group. In the ¹H-NMR spectrum, four sets of protons were appeared at $\delta_{\rm H}$ 4.32 (2H, dd, J= 11.7, 3.0 Hz, H- 1 A and H-6 A), 4.12 (2H, d, J= 7.1 Hz, H-3 and H-4), 3.60 (2H, m, H-2 and H-5) and 3.50 (2H, dd, J= 11.7 and 6.0 Hz, H-1_B and H-6_B).

Compound **10** was identified as galactitol by comparing its physical and spectral composition with the literature data.^{29,30} This is the first isolation of galactitol from the genus *Ixora*.

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